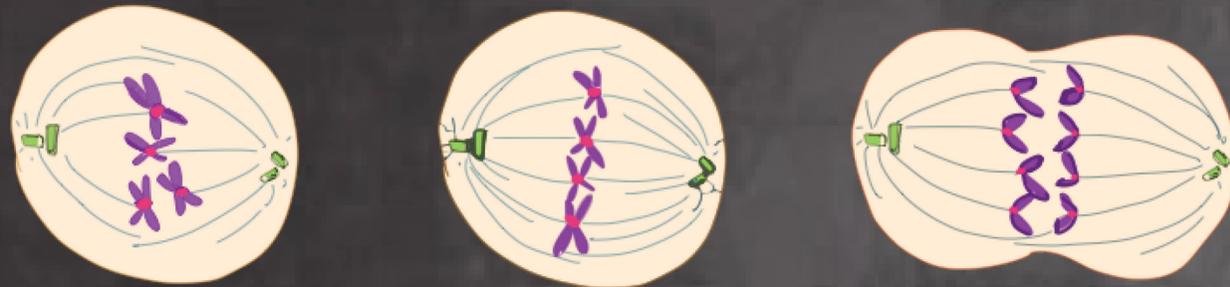


Assembly principles and stoichiometry of a complete human kinetochore module

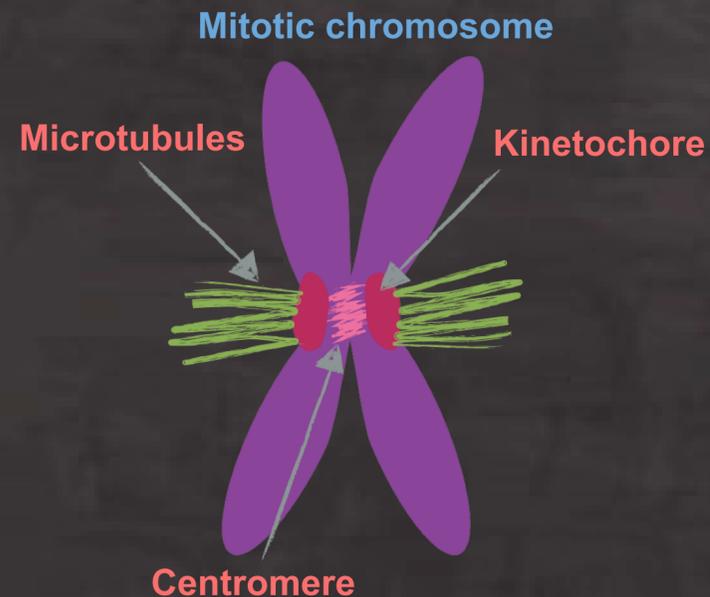
Science Advances, Vol. 7, no. 27, 2021.

Introduction

The Musacchio lab aims at understanding the molecular components and processes involved in eukaryotic cell division. The distribution of two exact copies of the genome from the mother to the two daughter cells depends on a large number of protein interactions and regulation systems.



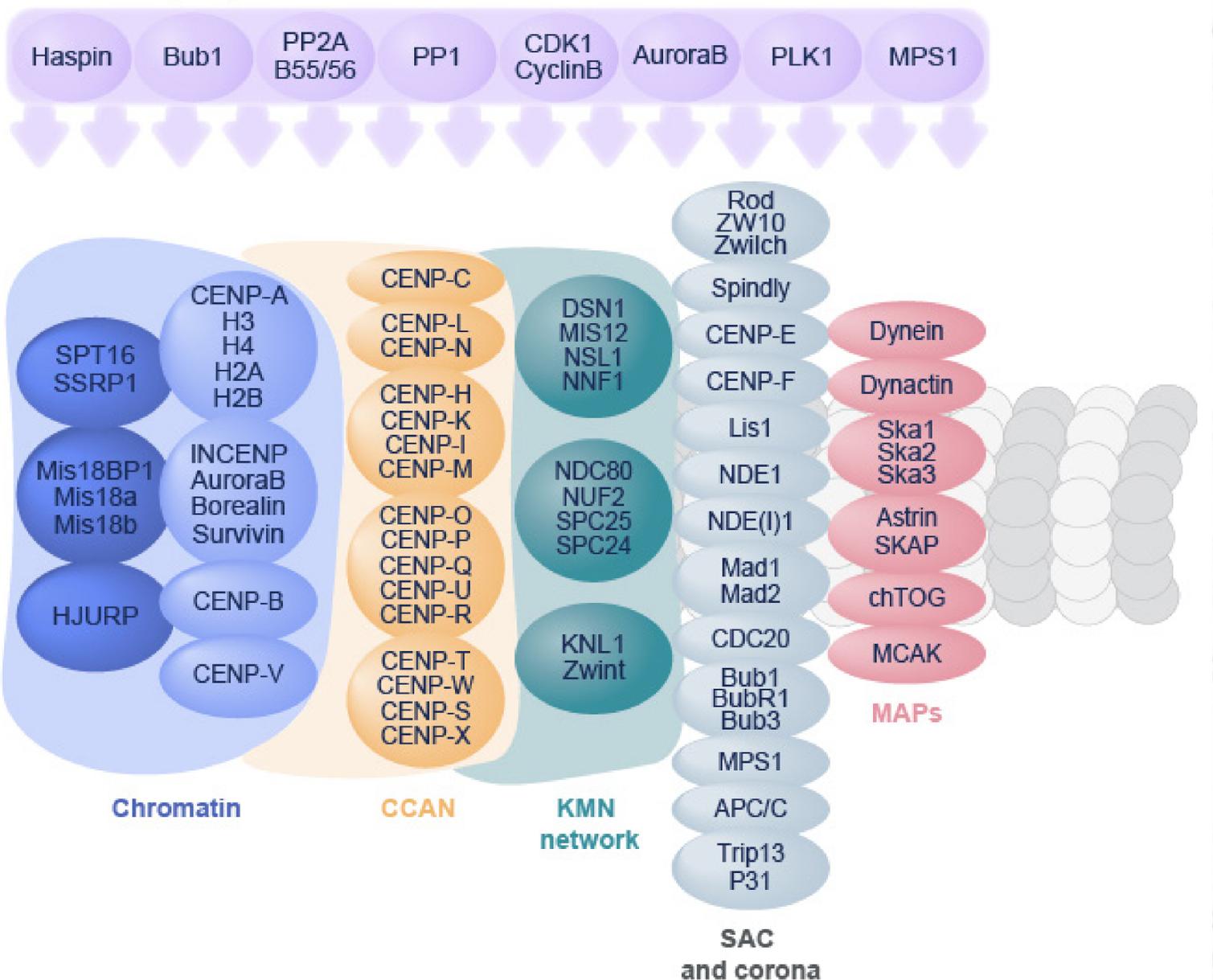
For the physical separation of the two sister chromatids, the centromere is a key player. It is a specific chromosomal region that binds both chromatids together and enables kinetochore assembly and microtubule attachment.



In this study we look closer at how centromeric chromatin links to kinetochore assembly and centromere identity. For this, we reconstituted a minimal, simplified kinetochore module containing all known core kinetochore subunits at stoichiometries that resemble those of previously reported intracellular quantifications of entire human kinetochores.

Recombinant kinetochore in Musacchio Lab

Kinases and phosphatases



Main Players

CENP-C:

- is the largest CCAN member and provides a blueprint for kinetochore assembly
- contains an N-terminal binding site for the MIS12 complex, two conserved motifs mediating binding to CENP-A nucleosomes, a binding region for the CCAN members CENP-HIKM/-LN and a C-terminal dimerization domain
- is crucial for the assembly of a complete kinetochore

CENP-T:

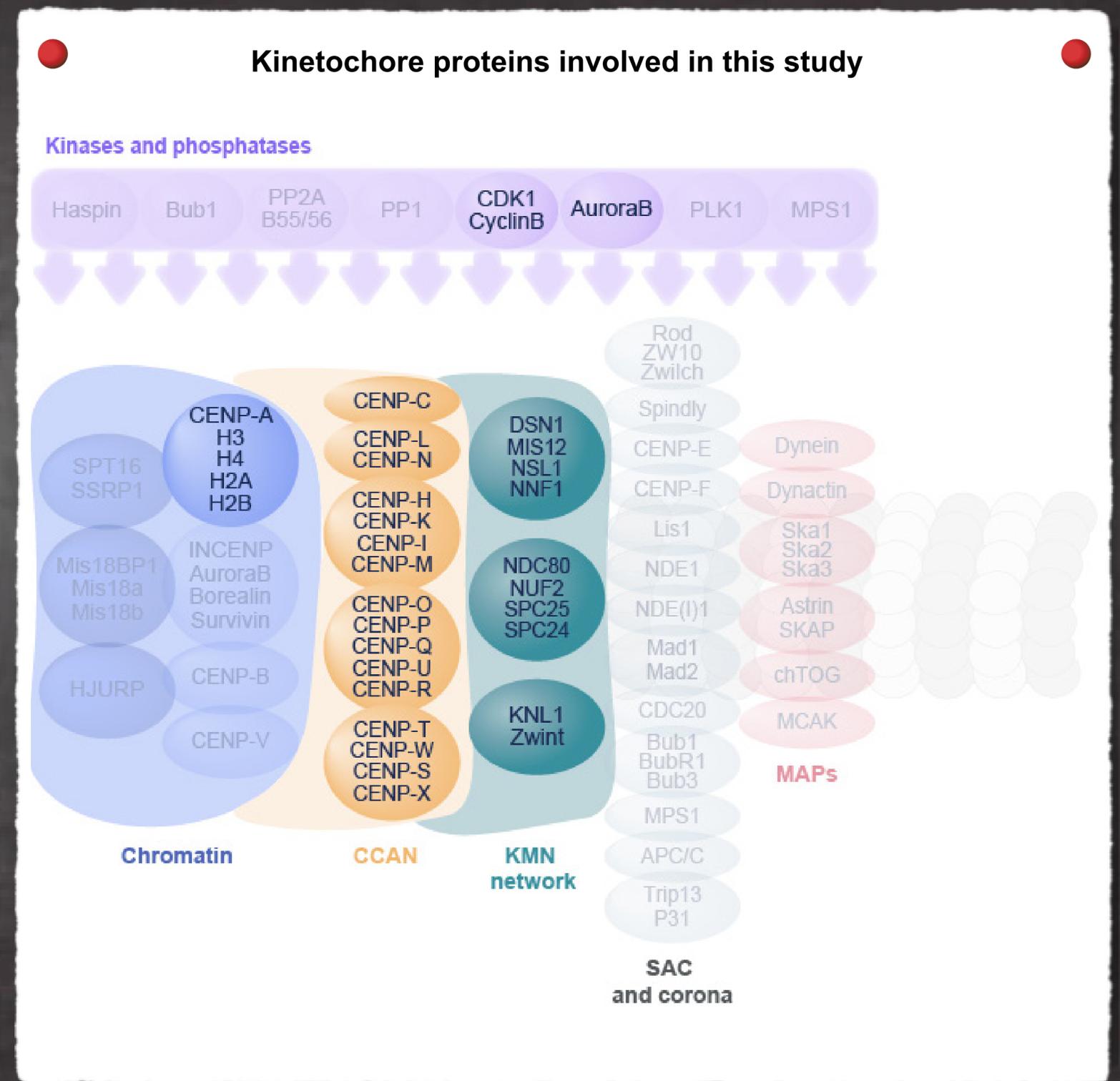
- forms a link to the outer kinetochore besides CENP-C
- recruits two NDC80 complexes and one KMN network in a CDK1-Cyclin B phosphorylation dependent manner

CENP-A and H3 nucleosomes:

- Nucleosomes are the fundamental subunit of the chromatin
- consist of a segment of DNA that is wrapped around a histone octamer
- CENP-A is a histone H3 variant specifically enriched at the centromere
- CENP-A is recognized by two CCAN members, CENP-N and CENP-C

Aurora B kinase:

- phosphorylates various kinetochore proteins
- enables the binding between the MIS12 complex and CENP-C



Questions

Can we reconstitute a simplified, but **complete kinetochore module** that represents the minimal building block of the human kinetochore?

How many nucleosomes are included in such a kinetochore module?

Can we incorporate the outer kinetochore components at the **right stoichiometries**?

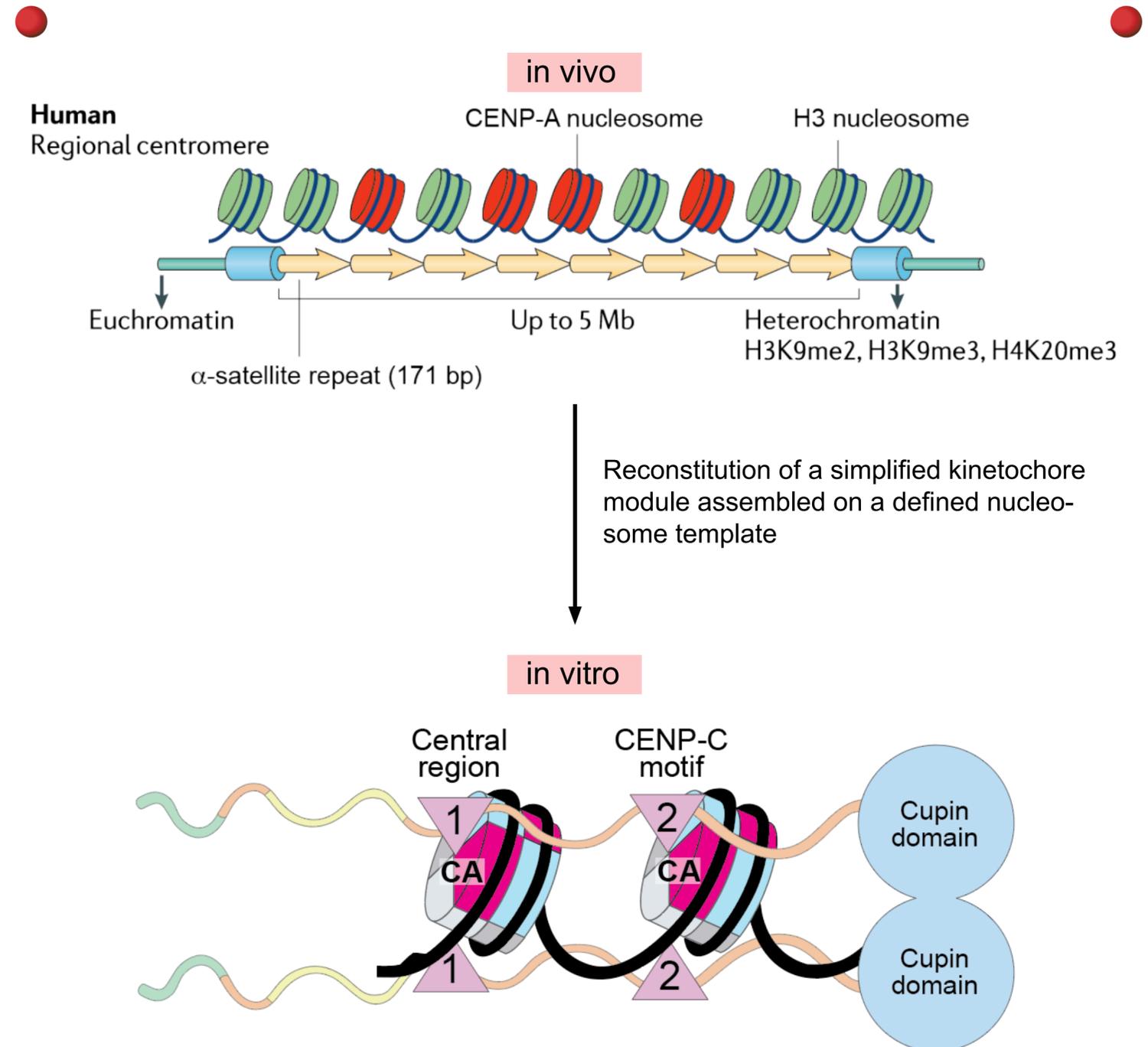
Can we reconstitute **full-length CENP-C**, a key component for kinetochore assembly, and demonstrate that it is functional?

Which **binding sites of CENP-C** are crucial for its centromere targeting?

What is the role of **CENP-C dimerization**?

How is the **CENP-T:MIS12** complex interaction regulated?

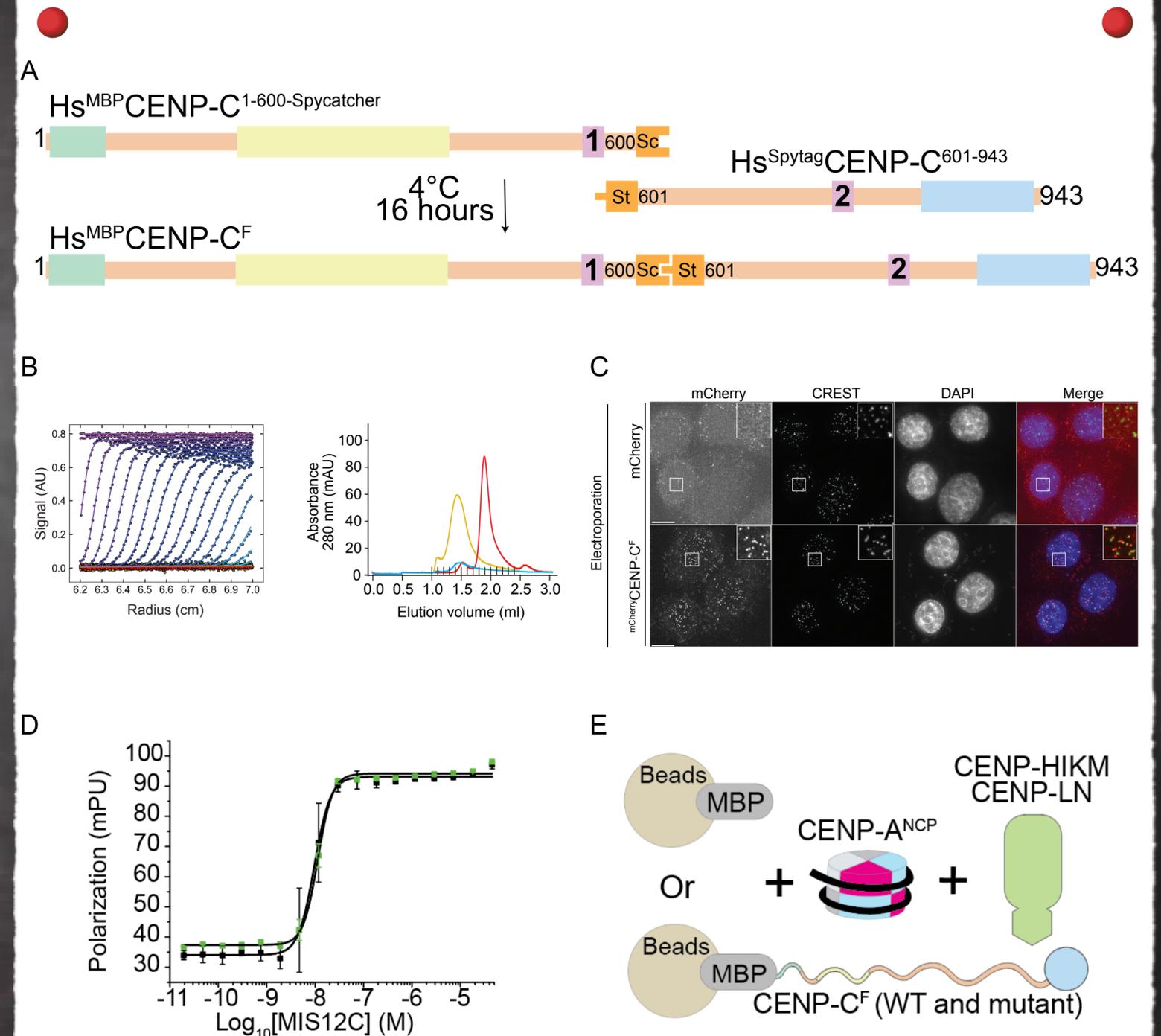
Can we explain why one **MIS12** complex cannot simultaneously bind to CENP-C and CENP-T?



Main methods used

In this study we used

-  The **SpyCatcher/SpyTag** system to obtain a full-length version of human CENP-C by joining together two complementary CENP-C fragments (A)
-  Powerful biophysical methods like **Analytical Ultracentrifugation** and **size-exclusion chromatography** to determine the molecular weight and stoichiometries of complexes containing full-length dimeric CENP-C mixed with CENP-A nucleosomes at defined ratios (B)
-  **Electroporation** as a tool for the efficient and rapid delivery of recombinant fluorescently tagged proteins into living human cells to test the subcellular localization and the functionality of wild type CENP-C and CENP-T as well as site directed mutants (C)
-  **Fluorescence polarization assays** to identify residues of CENP-T that are critical for the interaction with the MIS12 complex (D)
-  **Pull-down assays** on amylose resin to reconstitute large megadalton-scale complexes containing all main components of the human kinetochore on solid phase (E)



Results

💡 We reconstituted a full-length **CENP-C** and demonstrated that cells with electroporated **mCherry-CENP-C** were functional at the same level as cells with endogenous **CENP-C**. (A)

💡 We discovered that the **CENP-HIKM/-LN** binding sites are crucial for the centromere recruitment of **CENP-C**.

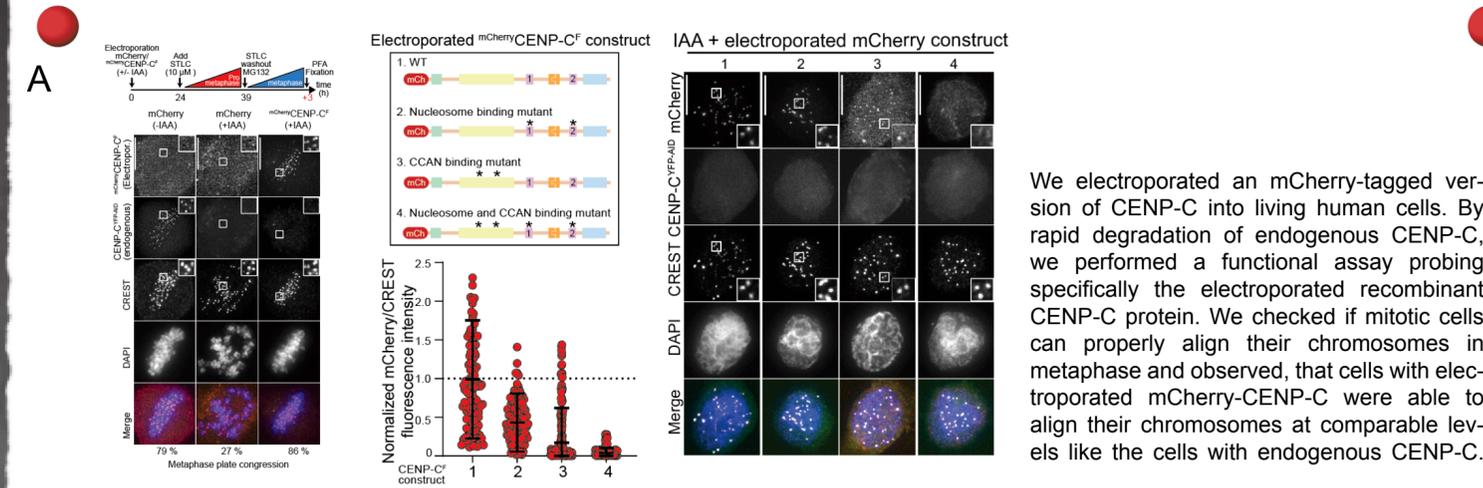
💡 We determined that the dimerization of the Cupin domain stabilizes kinetochore **CENP-C**, probably through enforcement of multivalent interactions with other CCAN subunits.

💡 We showed that for the **CENP-T:MIS12** complex interaction, **CENP-T** needs to be phosphorylated by **CDK1**, and the **MIS12** complex needs to be phosphorylated by **Aurora B**.

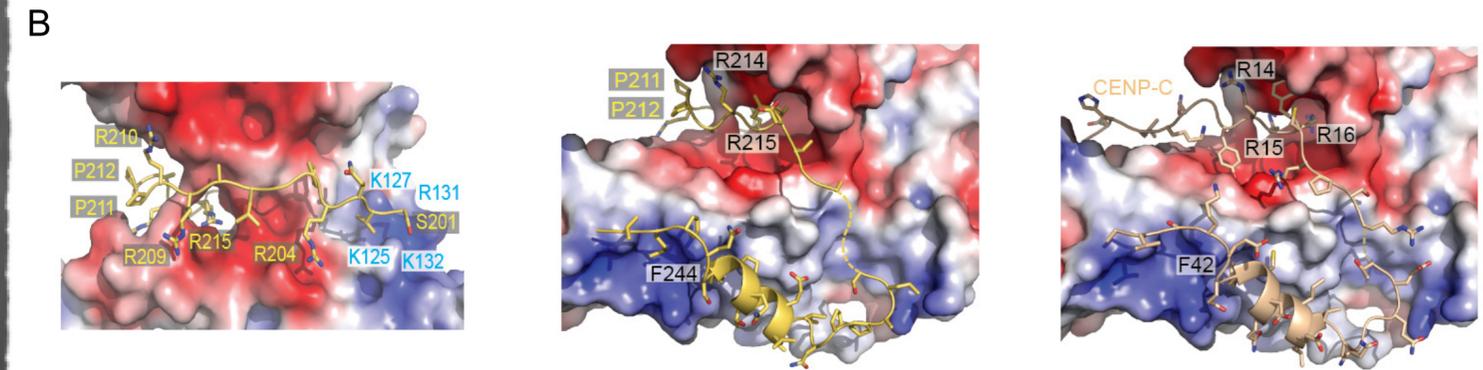
💡 We created a model describing the competitive binding of **CENP-C** and **CENP-T** to the **MIS12** complex. (B)

For the first time, we reconstituted a complete kinetochore containing all core components incorporated with the expected stoichiometries. Our final model summarizes the obtained results:

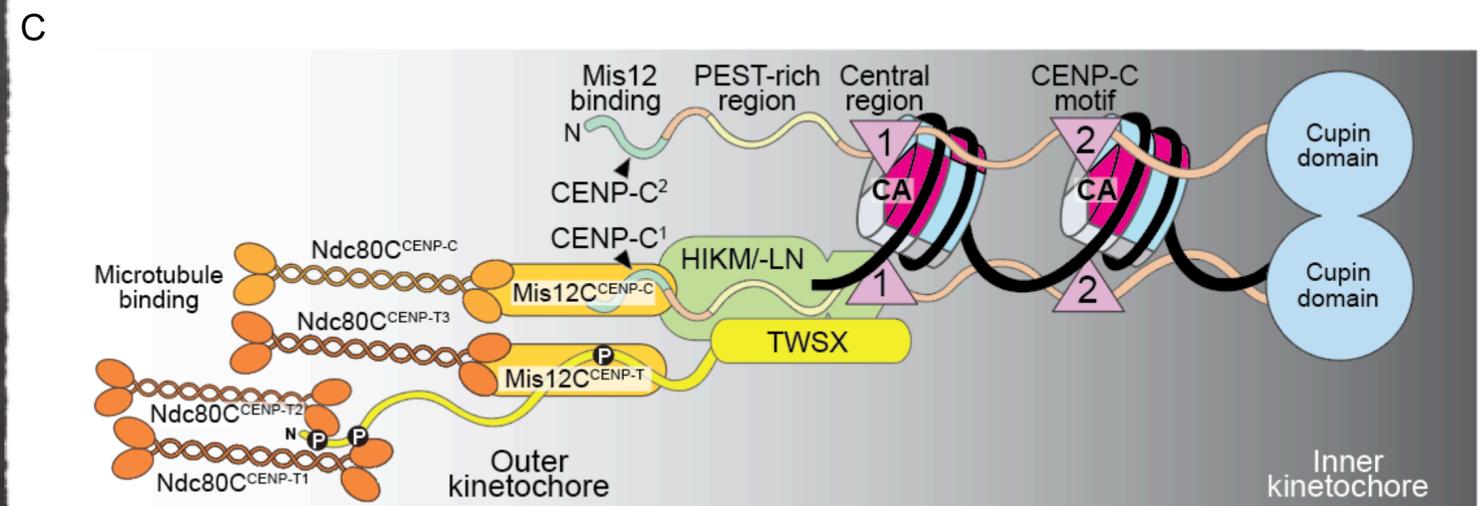
The **CENP-C** dimer binds two **CENP-A** nucleosomes and two copies of **CENP-OPQR** (not depicted), **CENP-LN** and **CENP-HIKM**, the latter recruits **CENP-TWSX** which binds two **NDC80** complexes and one additional **KMN** complex (containing **KNL1**, the **MIS12** complex and the **NDC80** complex). The **CENP-C** N-Terminus binds another **KMN** complex. (C)



We electroporated an mCherry-tagged version of CENP-C into living human cells. By rapid degradation of endogenous CENP-C, we performed a functional assay probing specifically the electroporated recombinant CENP-C protein. We checked if mitotic cells can properly align their chromosomes in metaphase and observed, that cells with electroporated mCherry-CENP-C were able to align their chromosomes at comparable levels like the cells with endogenous CENP-C.



With the help of Crosslink-MS analyses, we modeled the binding of a short CENP-T peptide to the MIS12 complex. The first part of the CENP-T peptide occupies a unique ridge in the MIS12 complex structure (left) until it bends to interact with the previously described CENP-C interface (middle and right).



Outlook

By combining biochemical reconstitution and structural biology, we want to gain detailed insights into the organization of centromeric nucleosomes and their interaction with kinetochore proteins.

Acknowledgements

We are grateful to **Daniele Fachinetti** and **Don C. Cleveland** for sharing reagents.

To **Ingrid Hoffmann**, **Carolin Koerner**, **Lisa Schulze**, **Isabelle Stender**, **Annika Take**, **Beate Voss**, and **Sabine Wohlgemuth** for general technical assistance, and to **Franziska Müller** and **Petra Janning** for help with mass spectrometry experiments.

AM gratefully acknowledges funding by the **Max Planck Society**, the **European Research Council (ERC) Advanced Investigator Grant RECEPIANCE** (proposal 669686), and the **DFG's Collaborative Research Centre (CRC) 1093**.



Kai Walstein



Andrea Musacchio



Dongqing Pan



Ingrid Vetter



Arsen Petrovic



Doro Vogt



Birte Hagemeyer